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# Two High-Nutrient Low-Chlorophyll phytoplankton assemblages: the tropical central Pacific and the offshore Perú-Chile Current

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**Abstract.** The phytoplankton ( $>15\ \mu\text{m}$ ) composition and abundance was investigated along a  $\sim 8000\text{ km}$  transect between the Marquesas Islands Archipelago and the Chilean coasts off Concepción. In the southern limit of the central Equatorial Pacific (at  $8^\circ\text{ S}$ ,  $141^\circ\text{ W}$ ), in High-Nutrient Low-Chlorophyll (HNLC) warm waters, the microphytoplankton assemblage was dominated by the lightly silicified diatoms *Pseudo-nitzschia delicatissima* and *Rhizosolenia bergonii*. The morphology of these species, a small pennate diatom that exhibited a tendency to form “ball of needles” clusters and large centric diatom ( $>500\ \mu\text{m}$  long), are interpreted as two anti-grazing strategies in an environment dominated by small micrograzers. Surprisingly, this a priori typical HNLC phytoplankton assemblage was also found in the temperate offshore waters of the Perú-Chile Current between 2000 and 600 km off Chile. This observation suggests that a common set of environmental factors (obviously other than temperature and salinity) are responsible for the establishment and maintaining of this distinctive phytoplankton in these geographically and hydrologically distant regions. Both regions are characterized by a surface nitrate-silicic acid ratio ranging from 1–3. Occasionally *Rhizosolenia bergonii* showed frustules anomalously fragmented, likely the result of extreme weakly silicified phytoplankton. We suggest that silicon deficiency may be responsible of the occurrence of HNLC phytoplankton assemblage in the tropical central Pacific as well as the offshore Perú-Chile Current during the austral summer.

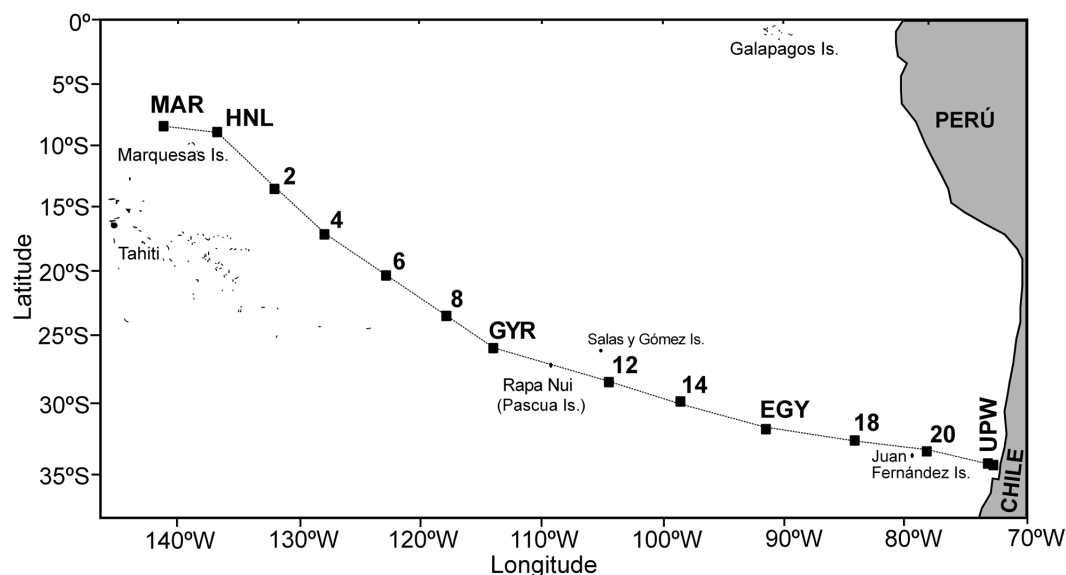
## 1 Introduction

The equatorial upwelling region in the central and eastern Pacific constitutes a large area of the so-called the High Nutrient Low Chlorophyll (HNLC) waters, representing a strong source of  $\text{CO}_2$  to the atmosphere (Chavez and Toggweiler, 1995). Grazing control, ammonium inhibition of nitrogen uptake, limitations in silicic acid, micronutrient (iron) or light, lack of neritic bloom-forming diatoms among other hypothesis have been invoked to explain these paradoxical HNLC conditions (Chavez et al., 1991; Coale et al., 1996; Dugdale and Wilkerson, 1998). The microscope observations in the upwelling region of the central and eastern equatorial Pacific revealed a recurrent phytoplankton assemblage mainly composed of the small pennate diatom *Pseudo-nitzschia delicatissima* and the large centric diatom *Rhizosolenia bergonii* (Hasle, 1960).

The iron-hypothesis has challenged new studies in the equatorial upwelling region that have been focused, in particular, on biogeochemical fluxes and parameters such as biomass estimates and rate measures of primary and secondary production following the iron fertilization. These studies were nevertheless accompanied by scarce investigations on the large phytoplankton composition (Kaczmarek and Fryxell, 1994; Iriarte and Fryxell, 1995). They were often limited to identifications at suprageneric levels (Chavez and Buck, 1990) or focused on the species identification during iron-enriched incubation experiments (Fryxell and Kaczmarek, 1995).

The Perú-Chile or Humboldt Current originates when a part of the water that flows toward the east across the sub-Antarctic Ocean is deflected toward the north as it approaches South America. It flows northward along the coast

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**Fig. 1.** Map of the stations occupied (solid squares) in the SE Pacific Ocean.

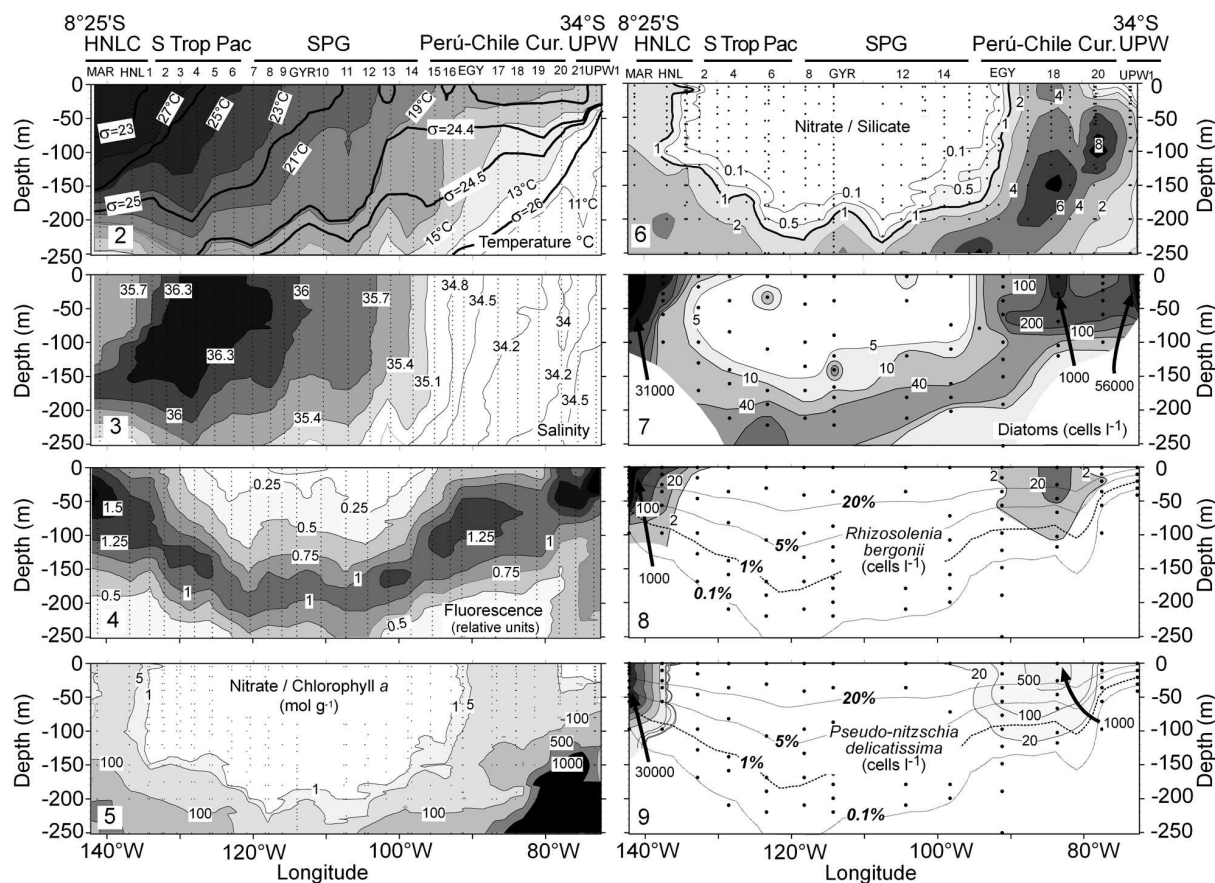
of Chile, Perú and Ecuador, and is associated with the economically most important coastal upwelling of the world's oceans (Longhurst, 1998). The coastal phytoplankton of the region is well documented (e.g. Rivera, 1983; Avaria and Muñoz, 1987). In contrast, the phytoplankton from the open waters of SE Pacific is one of the least known and documented of the world's oceans (Balech, 1962), reflecting the general undersampling of this remote area (Longhurst, 1998; Claustre and Maritorena, 2003). Very few recent investigations have been dedicated to the study of the picophytoplankton in the subtropical SE Pacific in latitudinal transects crossing the equator (DiTullio et al., 2003) or a few surface net samples with scarce detail on the phytoplankton specific composition (Hardy et al., 1996).

Within this context, we examined the composition and abundance of phytoplankton in relation to physical and chemical properties in the SE Pacific Ocean as part of a multidisciplinary survey along a transect of ~8000 km covering contrasted trophic conditions: HNLC waters of the Marquesas Islands Archipelago, hyper oligotrophic waters of the South Pacific Gyre (SPG), and more eutrophic conditions in the Perú-Chile Current (PCC) and the Chilean coastal upwelling. The present study focuses on the HNLC phytoplankton assemblage (HNLC-PA) near the Marquesas Islands Archipelago, the southern limit of the equatorial upwelling region that unexpectedly was also observed in a vast region of the temperate waters of the SE Pacific off Chile. The similarity in HNLC phytoplankton assemblage between two distant geographical and hydrological regions is analyzed and interpreted in the context of possible driving factors precluding this a priori paradoxical situation.

## 2 Methods

The data were collected during the BIOSOPE (Biogeochemistry and Optics South Pacific Experiment) cruise on board R/V L'Atalante from 26 October–12 December 2004. The ship was equipped with a 75 kHz RDI Acoustic Doppler Current Profiler (ADCP). Water sampling and measurements of temperature and salinity were made using a SeaBird SBE 911plus CTD/Carousel system fitted with an in situ Aquatracka III Chelsea fluorometer and Niskin bottles. The percentage of surface irradiance at each depth was calculated from the underwater PAR (Photosynthetic Active Radiation, 400–700 nm) profiles performed by a PNF-300 Profiling Natural Fluorometer sensor (Biospherical Instruments, Inc.). The limit of the euphotic zone corresponds to the depth where PAR is reduced to 1% of its surface value. Major nutrients and chlorophyll *a* were measured as described in Raimbault et al. (2007).

For microplankton analysis, samples were collected at 14 stations from 5–280 m depth (Fig. 1). One hundred samples were preserved with acidified Lugol's solution and stored at 5°C. A 500 ml aliquot of the sample was concentrated by sedimentation in glass cylinders. During a six-day settling period, the top 450 ml of the sample was slowly siphoned off with small-bore tubing. Fifty ml of the concentrate representing 500 ml of whole water sample was used for settlement in a composite settling chamber. The entire chamber was scanned at 200× magnification under an Olympus inverted microscope equipped with a digital camera. The specimens were photographed at 400× magnification with the Olympus DP70-BSW software. This methodology is suitable for the counting and identification of organisms higher than 15 µm. Diatoms and dinoflagellates were



**Fig. 2–9.** Section plots of (2) temperature ( $^{\circ}\text{C}$ ) and the isopycnals lines labelled in units of sigma- $t$  (density-1000)( $\text{kg m}^{-3}$ ) are shown. (3) Salinity. (4) Fluorescence (relative units). (5) Nitrate-chlorophyll  $a$  ratio. (6) Nitrate-silicic acid ratio. (7) Abundance of diatoms. (8) *Rhizosolenia bergonii* and (9) *Pseudo-nitzschia delicatissima* ( $\text{cells l}^{-1}$ ). The dashed lines represent the percentage of the surface irradiance.

well preserved, while most nanoplankton was lost due to preservation (i.e. coccolithophorids) or incomplete sedimentation. The organisms were identified to species level when possible.

### 3 Results

#### 3.1 Tropical HNLC Phytoplankton assemblage

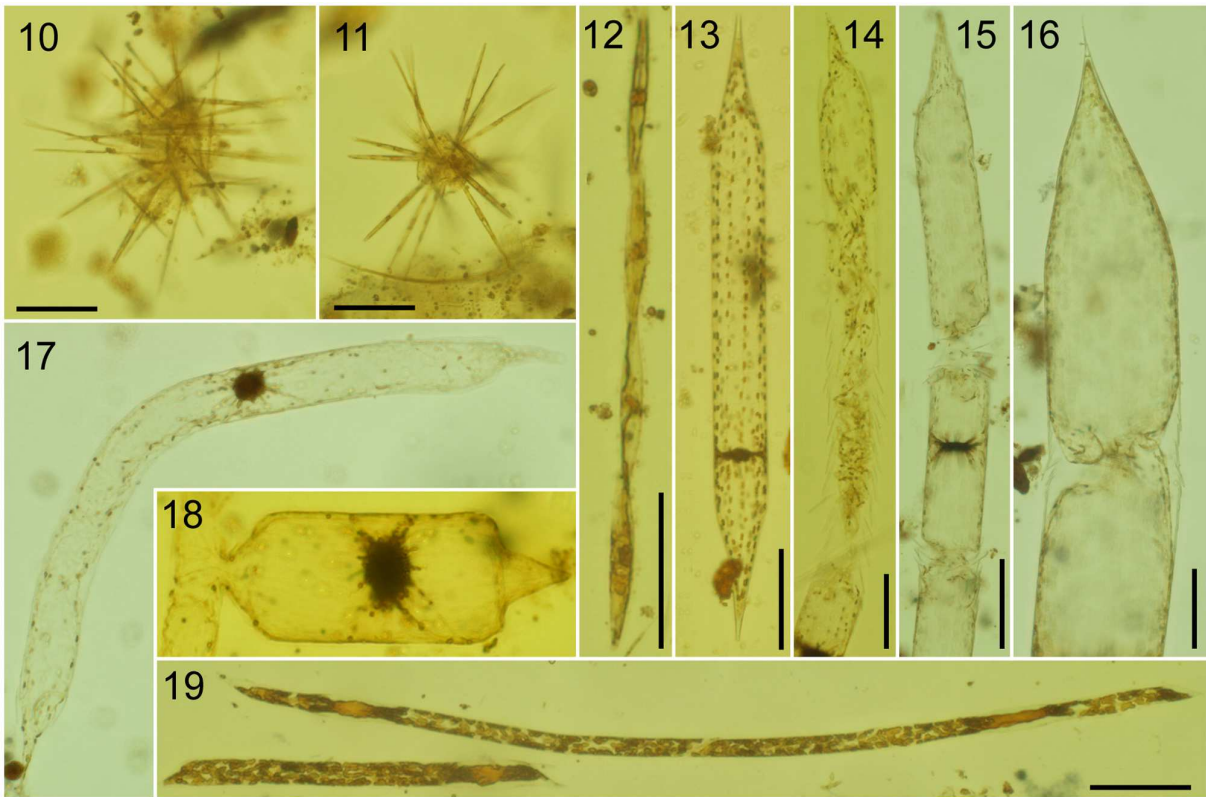
Waters surrounding the Marquesas Islands ( $8^{\circ}\text{S}$ ,  $141^{\circ}\text{W}$ ) under the influence of the South Equatorial Current constituted the southern border of HNLC waters of the Equatorial Upwelling Region. Salinity ranged from 35.5–35.7 and the surface temperature reached  $27.9^{\circ}\text{C}$ . The vertical distribution of the fluorescence showed a wide maximum in the upper 50 m depth (Figs. 2–4). The concentrations of chlorophyll  $a$  (Chl  $a$ ) were about  $0.3\text{--}0.5\text{ }\mu\text{g l}^{-1}$  with values of the nitrate-Chl  $a$  ratio higher than 5 (Fig. 5, Table 1).

Along 600 km between  $8^{\circ}23'\text{S}$  and  $13^{\circ}32'\text{S}$  extended the tropical HNLC-PA mainly represented small pennate diatom *Pseudo-nitzschia delicatissima* ( $30\text{--}80\text{ }\mu\text{m}$  in length,

$2\text{--}3\text{ }\mu\text{m}$  in width) and the large centric diatom *Rhizosolenia bergonii* ( $>500\text{ }\mu\text{m}$  in length). The microphytoplankton abundance reached up to  $31\,000\text{ cells l}^{-1}$  in the upper 60 m depth of the surrounding waters the Marquesas Islands Archipelago (Fig. 7). *Pseudo-nitzschia delicatissima* dominated the HNLC-PA with more than 85% of the total abundance. This threadlike and lightly sicilified species was relatively homogeneously distributed in the upper 60 m depth, and often forming clusters of 10–20 cells (Figs. 10–11). Other abundant colonial congeneric species were *Pseudo-nitzschia* cf. *subpacific* (Fig. 12) and *Pseudo-nitzschia* cf. *pungens*. Strongly elongated cells (up to  $2000\text{ }\mu\text{m}$  in length) of the pennate diatom *Thalassiothrix longissima* reached densities of  $1200\text{ cells l}^{-1}$  and other diatoms such as *Plagiotropis* spp. and *Haslea* spp. were abundant. *Rhizosolenia bergonii* was dominant and reached an abundance of  $1000\text{ cells l}^{-1}$  (maximum at 40–60 m depth, Figs. 13–14), followed by *Rhizosolenia acuminata* ( $120\text{ cells l}^{-1}$ ). Although the abundance of these diatoms was low compared to *Pseudo-nitzschia* spp., it should be taken into account that they are very large and thick, and contribute importantly to

**Table 1.** Average values in the euphotic zone of chlorophyll *a* (Chl *a*,  $\mu\text{g l}^{-1}$ ), nitrate ( $\text{NO}_3^-$ ,  $\mu\text{M}$ ), phosphate ( $\text{PO}_4^{3-}$ ,  $\mu\text{M}$ ), silicic acid ( $\text{Si(OH)}_4$ ,  $\mu\text{M}$ ), nitrate-silicic acid ratio (N/Si), nitrate-phosphate ratio (N/P), abundance of diatoms ( $> 15 \mu\text{m}$ ) and dinoflagellates ( $> 15 \mu\text{m}$ ), units expressed as cells  $\text{l}^{-1}$ . Ze=depth of the euphotic zone calculated from the PNF-300 Profiling Natural Fluorometer sensor.

HNLC-Marquesas			Tropical and South Pacific Gyre							Perú-Chile Current			Upw.
Station	MAR1	HNL	2	4	6	8	GYR3	12	14	EGY3	18	20	UPW1
Ze	66	90	124	136	157	144	155	152	136	94	87	48	31
Chl <i>a</i>	0.46	0.24	0.17	0.12	0.03	0.07	0.10	0.01	0.09	0.19	0.20	0.53	1.10
$\text{NO}_3^-$	1.48	1.52	0	0.04	0.02	0	0.01	0.01	0.01	0.30	4.08	2.18	2.38
$\text{PO}_4^{3-}$	0.46	0.36	0.17	0.14	0.12	0.13	0.15	0.15	0.05	0.16	0.42	0.41	0.63
$\text{Si(OH)}_4$	1.17	1.27	0.59	1.04	1.15	1.16	0.85	0.86	0.83	0.99	0.57	0.68	2.56
N/Si	1.27	1.25	0	0.04	0.02	0	0.01	0.01	0.01	0.32	7.10	3.16	0.86
N/P	3.23	4.34	0	0.30	0.19	0	0.06	0.04	0.13	1.61	9.73	4.64	3.62
DIATO	25103	148	8	4	10	1	9	14	15	89	662	176	342548
DINO	117	140	53	34	30	23	34	36	39	49	71	805	783



**Fig. 10–19.** Photomicrographs of microphytoplankton in the SE Pacific. **(10–11)** Clusters of *Pseudo-nitzschia delicatissima*, 20 m and 30 m depth. **(12)** *Pseudo-nitzschia* cf. *subpacific*, 5 m depth. **(13)** “Normal” cell of *Rhizosolenia bergonii*, 60 m depth. **(14)** *R. bergonii* with fragmented frustule, 20 m depth. (10–14) 8°23' S; 141°14' W. **(15)** *R. bergonii* with fragmented frustule, 30 m depth. **(16)** *Rhizosolenia acuminata*, 5 m depth. (14–16) 9° S, 136°51' W; 30 and 5 m depth. **(17)** Curved cells of *R. bergonii* (13°32' S, 132°07' W, 5 m depth). **(18)** Auxospore of *R. bergonii*, 33°21' S, 78°06' W, 5 m depth. **(19)** Highly pigmented cells of *Rhizosolenia* sp. (17°13' S, 127°58' W, 210 m depth). Scale bar=50  $\mu\text{m}$ .

the phytoplankton biomass. Other common centric diatoms were *Chaetoceros peruvianus* (200 cells  $\text{l}^{-1}$  near surface), *Chaetoceros atlanticus* var. *neapolitanus*, *Planktoniella sol*, *Bacteriastrium* cf. *elongatum* and *Asteromphalus heptactis*.

The dinoflagellates, ranging from 112–180 cells  $\text{l}^{-1}$ , showed a shallower distribution than the diatoms. The assemblage was rich on large autotrophic thecate species of the genus *Ceratium* (48–108 cells  $\text{l}^{-1}$ ), mainly *C. tripos*, *C. furca*,

*C. massiliense*, *C. candelabrum*, and unarmoured dinoflagellates such as *Brachidinium capitatum* f. *Karenia papilionacea* (12–24 cells  $l^{-1}$ ), *Pyrocystis* spp. and the prasinophyte in phycome stage *Pterosperma moebii* (40 cells  $l^{-1}$ ). Among the heterotrophic species, dinoflagellates (*Protoperidinium* and *Gyrodinium*) and choanoflagellates were abundant (Table 2).

At a station located 480 km eastwards (9° S, 136° 51' W), diatom abundance decreased to 100–380 cells  $l^{-1}$  (Fig. 7) and was dominated by *Rhizosolenia bergonii* that accounted for half of diatoms (Fig. 8). The abundance of *Pseudo-nitzschia delicatissima* was low (<140 cells  $l^{-1}$ ) and the diatom clusters were not observed. The abundance of *R. bergonii* (180 cells  $l^{-1}$ ) was less reduced than that of *P. delicatissima* in proportion to the previous station (Fig. 9). As occurred in the previous station, specimens of *R. bergonii* and *R. acuminata* showed the frustule anomalously fragmented in some concentric sections of the girdle (Figs. 14–16).

The decrease of the diatom abundance was associated with an increase of dinoflagellates (100–230 cells  $l^{-1}$ ). The autotrophic species of *Ceratium* were replaced by heterotrophic species such as *C. teres*. The abundance of diatom-consumers dinoflagellates (*Protoperidinium* spp.) decreased and smaller heterotrophic species such as *Oxytoxum variable* and *Podolompas spinifera* were common.

To the south (13° 32' S, 132° 07' W), the HNLC-PA was present with a very low abundance (<50 cells  $l^{-1}$ ) (Figs. 7–9). *Pseudo-nitzschia delicatissima* (20 cells  $l^{-1}$ ), *Planktoniella sol*, a few specimens of *R. bergonii*, and *Thalassiothrix* sp. were encountered in the surface layer. Near the bottom of the euphotic (100–150 m depth, see Fig. 3), the salty South Tropical Pacific Waters coincided with highly pigmented specimens of *Rhizosolenia* cf. *imbricata* (Fig. 19), and *Nitzschia bicapitata* species complex at 170 m depth. The abundance of dinoflagellates ranged from 40–110 cells  $l^{-1}$ .

### 3.2 Temperate HNLC phytoplankton assemblage

Between 30° S and 32° S extended a subtropical front characterized by decrease of the salinity and temperature, and the shoaling of the isopycne of 1025.4  $Kg\ m^{-3}$ . The isohalines between 35.3 and 34.6 were vertically distributed in the upper 200 m. From 50–200 m depth, the isotherms were vertically distributed decreasing between 18°C and 16°C (Figs. 2–3). To the east, along the salinity frontal region the fluorescence increased and shoaled between 120 m and 70–80 m depth (Fig. 4). The nitrate-Chl *a* ratio showed values higher than 5 (Fig. 5).

The Perú-Chile Current is a width equatorward cold-water current. The isopycne of 1025.4  $kg\ m^{-3}$  (Fig. 2) and the depth of the fluorescence maximum were horizontally distributed (Fig. 4). The PCC, delimited by isohaline of 34.3, was associated with fresher waters, especially between

78° W and 75° W (salinity  $\sim 34.1$ ). Near the Juan Fernández Archipelago (650 km off Chile) the salinity decreased to 33.9 and isotherm of 11°C shoaled (Figs. 2–3). This fresher water mass found in the surface may correspond to the Eastern South Pacific Intermediate Water as defined by Schneider et al. (2003). In these conditions, the fluorescence increased and shoaled (Fig. 4). The ADCP measurements showed a northwards component between 92° W and 84° W, while the current was weak and often reverse towards the coasts of Chile.

Large phytoplankton was nearly absent in the upper 120 m of the vast region of the SPG with very deep maximum composed of *Nitzschia bicapitata* species complex, *Nitzschia* spp. and other small pennate diatoms. The large phytoplankton in the upper layer reappeared towards the western Perú-Chile Current. The surface phytoplankton assemblage found in the PCC coincided to that observed in the central tropical Pacific. About 2000 km off Chile under the influence of PCC (31° 52' S, 91° 24' W) the phytoplankton was constituted of the typical HNLC-PA in surface and deeper the diatom assemblage found in the oligotrophic waters of the SPG. In upper 100 m, the diatom abundance was 50–196 cells  $l^{-1}$  composed of *Pseudo-nitzschia delicatissima* (160 cells  $l^{-1}$ ), *Rhizosolenia bergonii* (8 cells  $l^{-1}$ ), *R. acuminata* (6 cells  $l^{-1}$ ), and a few specimens of *Plagiotropis* sp. and *Thalassiothrix* sp. Deeper than 100 m depth and parallel to the shoaling of the bottom of the euphotic zone, the abundance of the diatoms was low and composed of *Dactyliosolen* sp. (80–150 m depth), *Chaetoceros atlanticus* var. *neapolitanus* and *Thalassiosira* sp. (160 cells  $l^{-1}$  at 80 m depth) (Fig. 7).

In the upper 60 m depth, the abundance of dinoflagellates ranged from 54–168 cells  $l^{-1}$ . The autotrophic dinoflagellates *Gonyaulax polygramma* (60 cells  $l^{-1}$ ), *Ceratium fusus* (40 cells  $l^{-1}$ ), *C. furca* were responsible of the surface maximum. Other autotrophic species were *C. tripos*, *C. azoricum*, *C. candelabrum*, *C. kofoidii* as observed in the warm HNLC region and brachidiniaceans. A diverse assemblage of heterotrophic dinoflagellates of more eutrophic conditions was present (Table 2).

In temperate waters  $\sim 1300$  km off Chile (32° 42' S, 84° 04' W) the nitrate-silicic acid ratio was higher than 4 in the euphotic zone, with a maximum of 6 in the surface (Fig. 6, Table 1). The diatom assemblage was similar to that in HNLC conditions near the Marquesas Islands Archipelago. The diatom abundance showed values up to 1200 cells  $l^{-1}$  in upper layer (Fig. 7) dominated by *Pseudo-nitzschia delicatissima* (900 cells  $l^{-1}$ ), *Pseudo-nitzschia* cf. *subpacific* (120 cells  $l^{-1}$ ), *Rhizosolenia bergonii* (50 cells  $l^{-1}$ ) (Figs. 17–18), *Thalassiothrix* sp. (40 cells  $l^{-1}$ ), *Planktoniella sol* (16 cells  $l^{-1}$ ) and *Asteromphalus heptactis* (20 cells  $l^{-1}$ ). *Dactyliosolen* sp. and *Chaetoceros atlanticus* var. *neapolitanus* were encountered below the surface phytoplankton assemblage. The abundance of dinoflagellates was low (40–78 cells  $l^{-1}$ ), mainly composed of *Pronoctiluca pelagica* f. *spinifera*

**Table 2.** List of diatoms, dinoflagellates (> 15  $\mu\text{m}$ ) and large flagellates during the BIOSOPE cruise. HNLC=High-Nutrient Low-Chlorophyll region near the Marquesas Islands Archipelago; SPG=Tropical and South Pacific Gyre; PCC=Perú-Chile Current; UPW=Coastal upwelling off Concepción.

Diatom taxa	HNLC	SPG	PCC	UPW
<i>Asterionellopsis glacialis</i> (Castracane) Round				+
<i>Asterolampra marylandica</i> Ehrenberg		+		
<i>Asteromphalus flabellatus</i> (Brébisson) Greville	+	+	+	+
<i>Bacillaria</i> cf. <i>paxillifera</i> (O.F. Müller) Hendey	+			
<i>Bacteriastrum</i> cf. <i>comosum</i> Pavillard	+		+	
<i>Bacteriastrum</i> cf. <i>furcatum</i> Shadbolt	+			
<i>Bacteriastrum</i> sp1		+	+	
<i>Bacteriastrum</i> sp2				+
<i>Bacteriastrum</i> sp3		+		
<i>Chaetoceros affinis</i> Lauder				+
<i>Chaetocero atlanticus</i> var. <i>neapolitanus</i> (Schröder) Hustedt	+	+	+	
<i>Chaetoceros compressus</i> Lauder				+
<i>Chaetoceros curvisetus</i> Cleve				+
<i>Chaetoceros dadayi</i> Pavillard	+	+	+	
<i>Chaetoceros</i> cf. <i>diadema</i> (Ehrenberg) Gran				+
<i>Chaetoceros didymus</i> Ehrenberg				+
<i>Chaetoceros</i> cf. <i>diversus</i> Cleve		+		
<i>Chaetoceros laciniosus</i> Schütt				+
<i>Chaetoceros lauderi</i> Ralfs				+
<i>Chaetoceros lorenzianus</i> Grunow	+			+
<i>Chaetoceros peruvianus</i> Brightwell	+	+	+	
<i>Chaetoceros radicans</i> Schütt				+
<i>Chaetoceros rostratus</i> Lauder				+
<i>Chaetoceros</i> cf. <i>tenuissimus</i> Meunier				+
<i>Chaetoceros tetrastichon</i> Cleve	+	+		
<i>Chaetoceros</i> sp1		+		
<i>Chaetoceros</i> sp2		+		
<i>Chaetoceros</i> sp3	+	+		
<i>Chaetoceros</i> sp4		+		
<i>Chaetoceros</i> sp5		+		
<i>Corethron</i> cf. <i>criophilum</i> Castracane				+
<i>Coscinodiscus</i> sp.		+		
<i>Dactyliosolen</i> sp.		+	+	
<i>Dactyliosolen fragilissimus</i> (Bergon) Hasle				+
<i>Detonula pumila</i> (Castracane) Schütt				+
<i>Eucampia cornuta</i> (Cleve) Grunow				+
<i>Eucampia zodiacus</i> Ehrenberg				+
<i>Fragilariopsis doliolus</i> (Wallich) Medlin et Sims	+			
<i>Guinardia delicatula</i> (Cleve) Hasle				+
<i>Guinardia flaccida</i> (Castracane) Peragallo				+
<i>Guinardia striata</i> (Stolterfoth) Hasle				+
<i>Grossleriella tropica</i> Schütt		+		
<i>Hemiaulus hauckii</i> Grunow		+		
<i>Hemidiscus cuneiformis</i> Wallich	+			
<i>Lauderia annulata</i> Cleve				+
<i>Leptocylindrus danicus</i> Cleve				+
<i>Neostreptotheca torta</i> f. <i>triangularis</i> Stosch		+		
<i>Nitzschia bicapitata</i> species complex	+	+	+	
<i>Nitzschia longissima</i> (Brébisson) Ralfs	+		+	+

Table 2. Continued.

Diatom taxa	HNLC	SPG	PCC	UPW
<i>Nitzschia</i> sp. (? <i>braarudii</i> )		+	+	
<i>Odontella</i> cf. <i>longicrucis</i> (Greville) Hoban				+
<i>Plagiotropis</i> sp.	+	+	+	
<i>Plagiotropis</i> sp. (? <i>lepidoptera</i> )	+	+	+	
<i>Planktoniella</i> sol (Wallich) Schütt				
<i>Proboscia</i> alata (Brightwell) Sundström	+	+	+	+
<i>Pseudotriceratium cinnamomeum</i> (Greville) Grunow		+		
<i>Pseudo-nitzschia delicatissima</i> (Cleve) Heiden <i>sensu</i> Hasle 1960	+	+	+	
<i>Pseudo-nitzschia</i> cf. <i>pungens</i> (Grunow <i>ex</i> Cleve) Hasle	+		+	
<i>Pseudo-nitzschia</i> cf. <i>subpacifica</i> (Hasle) Hasle	+		+	+
<i>Pseudo-nitzschia</i> sp1		+		
<i>Pseudo-nitzschia</i> sp2				+
<i>Pseudo-nitzschia</i> sp3				+
cf. <i>Pseudoeunotia</i> sp.		+	+	
<i>Rhizosolenia acuminata</i> (Peragallo) Gran				
<i>Rhizosolenia bergonii</i> Peragallo	+	+	+	
<i>Rhizosolenia</i> cf. <i>castracanei</i> Peragallo		+		
<i>Rhizosolenia clevei</i> var. <i>communis</i> Sundström		+		
<i>Rhizosolenia</i> cf. <i>imbricata</i> Brightwell		+		+
<i>Rhizosolenia</i> cf. <i>imbricata</i> var. <i>shrubslei</i> (Cleve) Schröder		+		+
<i>Rhizosolenia setigera</i> Brightwell				+
<i>Rhizosolenia styliformis</i> Brightwell	+	+		
<i>Skeletonema</i> sp.				+
<i>Thalassionema nitzschioides</i> (Grunow) Mereschkowsky		+		+
<i>Thalassionema</i> cf. <i>pseudonitzschioides</i> (Schuette <i>et</i> Schrader) Hasle		+		+
<i>Thalassionema elegans</i> Hustedt	+	+		
<i>Thalassionema</i> sp.		+		
<i>Thalassiosira anguste-lineata</i> (A. Schmidt) Fryxell <i>et</i> Hasle				+
<i>Thalassiosira</i> cf. <i>decipiens</i> (Grunow) Jørgensen				+
<i>Thalassiosira</i> cf. <i>oestrupii</i> (Ostenfeld) Hasle	+		+	
<i>Thalassiosira subtilis</i> (Ostenfeld) Gran	+		+	+
<i>Thalassiosira</i> sp.			+	
<i>Thalassiothrix longissima</i> Cleve <i>et</i> Grunow	+	+	+	+
<i>Thalassiothrix</i> cf. <i>heteromorpha</i> Karsten	+	+	+	
Dinoflagellate taxa				
cf. <i>Amphidoma caudata</i> Halldal	+			
<i>Amphisolenia bidentata</i> Schröder		+	+	
<i>Amphisolenia globifera</i> Stein	+	+	+	
cf. <i>Blephaerocysta</i> sp.	+	+	+	
<i>Brachidinium capitatum</i> F.J.R. Taylor f. <i>Asterodinium</i>	+	+	+	+
<i>Brachidinium capitatum</i> F.J.R. Taylor f. <i>Brachidinium</i>	+	+	+	
<i>Brachidinium capitatum</i> F.J.R. Taylor f. <i>Karenia papilionacea</i>	+	+	+	+
<i>Brachidinium capitatum</i> F.J.R. Taylor f. <i>Microceratium</i>		+		
<i>Ceratium arietinum</i> Cleve		+		
<i>Ceratium azoricum</i> Cleve	+		+	+
<i>Ceratium belone</i> Cleve			+	
<i>Ceratium breve</i> (Ostenfeld <i>et</i> Schmidt) Schröder	+			
<i>Ceratium candelabrum</i> (Ehrenberg) Stein	+		+	
<i>Ceratium carnegiei</i> Graham <i>et</i> Bronikowsky	+		+	
<i>Ceratium carriense</i> Gourret	+	+	+	
<i>Ceratium</i> cf. <i>lineatum</i> (Ehrenberg) Cleve	+			



Table 2. Continued.

Dinoflagellate taxa	HNLC	SPG	PCC	UPW
<i>Ceratium contortum</i> (Gourret) Cleve			+	
<i>Ceratium declinatum</i> (Karsten) Jørgensen	+	+		
<i>Ceratium divaricatum</i> (Lemmermann) Kofoid var. <i>divaricatum</i>			+	
<i>Ceratium divaricatum</i> var. <i>balechii</i> (Meave, Okolodkov et Zamudio) Hernández-Becerril	+			+
<i>Ceratium extensum</i> (Gourret) Cleve		+	+	
<i>Ceratium falcatum</i> (Kofoid) Jørgensen	+			
<i>Ceratium furca</i> (Ehrenberg) Claparède et Lachmann	+	+	+	+
<i>Ceratium fusus</i> (Ehrenberg) Dujardin	+	+	+	+
<i>Ceratium gibberum</i> Gourret	+	+	+	
<i>Ceratium horridum</i> (Cleve) Gran				+
<i>Ceratium kofoidii</i> Jørgensen	+	+	+	
<i>Ceratium lanceolatum</i> Kofoid			+	
<i>Ceratium limulus</i> Gourret			+	
<i>Ceratium massiliense</i> (Gourret) Jørgensen	+	+	+	
<i>Ceratium pentagonum</i> Gourret	+	+	+	+
<i>Ceratium platycorne</i> Daday		+		
<i>Ceratium praeolongum</i> (Lemmermann) Kofoid			+	
<i>Ceratium pulchellum</i> Schröder				+
<i>Ceratium symmetricum</i> Pavillard		+	+	
<i>Ceratium teres</i> Kofoid	+	+		
<i>Ceratium trichoceros</i> (Ehrenberg) Kofoid	+	+	+	+
<i>Ceratium tripos</i> (O.F. Müller) Nitzsch	+	+	+	+
<i>Ceratocorys bipes</i> (Cleve) Kofoid			+	
<i>Ceratocorys horrida</i> Stein			+	
<i>Citharistes</i> sp.	+			
<i>Cladopyxis brachiolata</i> Stein	+	+	+	
<i>Cladopyxis</i> sp.	+	+	+	+
<i>Cochlodinium</i> sp.		+	+	+
<i>Craspedotella pileolus</i> Kofoid		+		
<i>Craspedotella</i> sp.	+	+	+	+
<i>Dicrionema psilonereia</i> F.J.R. Taylor et Cattell	+	+	+	
<i>Dinophysis acuminata</i> Claparède et Lachmann				+
<i>Dinophysis</i> cf. <i>swezyae</i> Kofoid et Skogsberg		+		
<i>Dinophysis fortii</i> Pavillard			+	
<i>Dinophysis mucronata</i> (Kofoid et Skogsberg) Sournia	+		+	
<i>Dinophysis schuettii</i> Murray et Whitting		+	+	
<i>Erythroplaxidium agile</i> (Hertwig) P.C. Silva	+	+	+	+
cf. <i>Gonyaulax birostris</i> Stein			+	
<i>Gonyaulax polygramma</i> Stein	+	+	+	
<i>Gymnodinium fusus</i> Schütt/Gyrodinium <i>falcatum</i> Kofoid et Swezy	+	+	+	
<i>Gymnodinium</i> sp1				+
<i>Gymnodinium</i> sp2		+		
<i>Gyrodinium</i> spp.	+			+
<i>Gyrodinium spirale</i> (Bergh) Kofoid et Swezy	+	+	+	+
<i>Histioneis</i> cf. <i>crateriformis</i> Stein		+	+	
<i>Histioneis</i> cf. <i>pulchra</i> Kofoid			+	
<i>Histioneis</i> cf. <i>striata</i> Kofoid et Michener			+	
<i>Histioneis cleaveri</i> Rampi	+	+		
<i>Histioneis hyalina</i> Kofoid et Michener	+			
<i>Histioneis joergensenii</i> Schiller		+		
<i>Histioneis longicollis</i> Kofoid			+	

Table 2. Continued.

Dinoflagellate taxa	HNLC	SPG	PCC	UPW
cf. <i>Histiophysis</i> sp.			+	
<i>Karenia</i> cf. <i>bicuneiformis</i> Botes, Sym et Pitcher		+		
<i>Karenia</i> cf. <i>mikimotoi</i> (Miyake et Kominami ex Oda) G. Hansen et Moestrup	+		+	
<i>Karenia</i> spp.	+	+	+	+
<i>Kofooidinium pavillardii</i> J. Cachon et M. Cachon	+	+	+	
<i>Kofooidinium</i> sp.	+	+	+	
<i>Kofooidinium velelloides</i> Pavillard	+	+	+	
<i>Leptodiscus medusoides</i> Hertwig			+	
cf. <i>Metaphalacroma skogsbergii</i> Tai in Tai et Skogsberg			+	
<i>Nematodinium</i> sp.	+	+	+	
<i>Ornithocercus magnificus</i> Stein		+	+	
<i>Ornithocercus quadratus</i> Schütt	+	+	+	
<i>Ornithocercus</i> sp.	+	+		
<i>Oxytoxum</i> cf. <i>crassum</i> Schiller	+	+	+	
<i>Oxytoxum</i> cf. <i>frenguelli</i> Rampi			+	
<i>Oxytoxum</i> cf. <i>laticeps</i> Schiller	+	+		+
<i>Oxytoxum</i> cf. <i>longum</i> Schiller		+	+	
<i>Oxytoxum challengeroides</i> Kofoid		+		
<i>Oxytoxum constrictum</i> (Stein) Bütschli		+	+	
<i>Oxytoxum curvatum</i> (Kofoid) Kofoid	+	+	+	
<i>Oxytoxum curvicaudatum</i> Kofoid		+	+	
<i>Oxytoxum diploconus</i> Stein	+	+	+	
<i>Oxytoxum longiceps</i> Schiller	+	+	+	
<i>Oxytoxum scolopax</i> Stein	+	+	+	+
<i>Oxytoxum</i> sp1			+	
<i>Oxytoxum</i> sp2	+			
<i>Oxytoxum</i> sp3		+		
<i>Oxytoxum</i> sp4			+	
<i>Oxytoxum tessellatum</i> (Stein) Schütt	+	+	+	
<i>Oxytoxum variabile</i> Schiller	+	+	+	
<i>Phalacroma rotundatum</i> (Claparède et Lachmann) Kofoid et Michener			+	
<i>Phalacroma</i> sp1	+	+	+	
<i>Phalacroma</i> sp2	+			
<i>Phalacroma</i> sp3		+		
<i>Phalacroma</i> sp4		+		
<i>Phalacroma</i> sp5		+	+	
<i>Podolampas bipes</i> Stein	+	+	+	
<i>Podolampas elegans</i> Schütt		+	+	+
<i>Podolampas palmipes</i> Stein	+	+	+	
<i>Podolampas</i> sp.	+			
<i>Podolampas spinifera</i> Okamura	+	+	+	+
<i>Polykrikos</i> cf. <i>schwartzii</i> Bütschli		+	+	+
cf. <i>Pomatodinium impatiens</i> J. Cachon et Cachon-Enjume	+		+	
<i>Pronoctiluca pelagica</i> Fabre-Domergue	+	+		
<i>Pronoctiluca pelagica</i> Fabre-Domergue f. <i>spinifera</i>	+	+	+	+
<i>Prorocentrum</i> cf. <i>compressum</i> (Bailey) Abé ex Dodge	+			
<i>Prorocentrum dentatum</i> Schiller	+	+		
<i>Prorocentrum rostratum</i> Stein	+	+	+	
<i>Prorocentrum</i> sp.			+	
<i>Protoperidinium</i> cf. <i>conicum</i> (Gran) Balech	+			
<i>Protoperidinium</i> cf. <i>curtipes</i> (Jørgensen) Balech	+		+	+

Table 2. Continued.

Dinoflagellate taxa	HNLC	SPG	PCC	UPW
<i>Protoperidinium</i> cf. <i>depressum</i> (Bailey) Balech				+
<i>Protoperidinium</i> cf. <i>divergens</i> (Ehrenberg) Balech	+			
<i>Protoperidinium</i> cf. <i>murrayi</i> (Kofoid) Hernández-Becerril	+		+	
<i>Protoperidinium</i> cf. <i>obtusum</i> (Karsten) Parke et Dodge	+			
<i>Protoperidinium</i> cf. <i>ovatum</i> Pouchet		+		
<i>Protoperidinium</i> cf. <i>steinii</i> (Jørgensen) Balech	+	+	+	+
<i>Protoperidinium</i> sp.	+			+
<i>Ptychodiscus noctiluca</i> Stein	+		+	
<i>Pyrocystis fusiformis</i> (Wyville Thomson ex Haeckel) Blackman	+			
<i>Pyrocystis lunula</i> (Schütt) Schütt	+	+	+	
<i>Pyrocystis noctiluca</i> J. Murray ex Haeckel	+	+	+	
cf. <i>Pyrocystis obtusa</i> Pavillard	+		+	
<i>Pyrophacus steinii</i> (Schiller) Wall et Dale			+	+
<i>Scaphodinium mirabile</i> Margalef	+	+	+	
<i>Scaphodinium</i> sp.		+		
<i>Scrippsiella</i> sp.		+		+
<i>Spatulodinium pseudonociluca</i> (Pouchet) J. Cachon et M. Cachon			+	+
<i>Spatulodinium</i> sp1		+	+	
<i>Spatulodinium</i> sp2		+		
cf. <i>Spiraulax jolliffei</i> (Murray et Whitting) Kofoid		+	+	
<i>Torodinium teredo</i> (Pouchet) Kofoid et Swezy	+	+	+	+
Large flagellate taxa				
<i>Dictyocha fibula</i> Ehrenberg	+		+	+
<i>Pterosperma moebii</i> (Jørgensen) Ostensfeld	+	+	+	
<i>Pterosperma cuboides</i> Gaarder		+		
<i>Halosphaera viridis</i> Schmitz		+		
<i>Solenicola setigera</i> Pavillard	+	+	+	+

(40 cells l<sup>-1</sup>), *Ptychodiscus noctiluca*, *Ornithocercus magnificus*, *Phalacroma rotundatum*, *Gyrodinium falcatum*, *Gonyaulax polygramma* (12 cells l<sup>-1</sup>), *Ceratium fusus*, and *Dinophysis fortii* (8 cells l<sup>-1</sup>) (Table 2).

About 650 km off Chile, a sampling station (33°21' S, 78°06' W) located 30 km north from the Robinson Crusoe Island, Juan Fernández Archipelago, was visited. The salinity reached a minimum of 33.9 in the surface. The ADCP measurements showed an eastwards drift and the surface warm waters run above the entrainment of fresher waters. This – surface stratified upwelling – resulted on a red tide of dinoflagellates with a surface abundance of 4400 cells l<sup>-1</sup>. The main component of the red tide was the non-toxic autotrophic dinoflagellate *Gonyaulax polygramma* that reached 3800 cells l<sup>-1</sup> at 5 m depth, 530 cells l<sup>-1</sup> at 25 m, and nearly disappeared below this depth. Other autotrophic dinoflagellates were *Ceratium azoricum* (90 cells l<sup>-1</sup>), *C. massiliense*, *C. gibberum*, *C. limulus*, *Ptychodiscus noctiluca* and brachidiniaceans. The heterotrophic dinoflagellates reached a high abundance, mainly composed of *Protoperidinium* cf. *curtipes* (120 cells l<sup>-1</sup>), *Ornithocercus quadratus*, *Gyrodinium spirale* in the upper 5–15 m depth as well as kofoidiniaceans (*Kofoidinium* and *Spatulodinium*). Dinoflagellates such as *Oxytoxum longum* and *Pronociluca*

*pelagica* f. *spinifera* were encountered at mid-depths. The abundance of diatoms ranged from 100–600 cells l<sup>-1</sup> (Fig. 7) with *Pseudo-nitzschia delicatissima*, *Rhizosolenia bergonii* (4 cells l<sup>-1</sup>), and *Pseudo-nitzschia* cf. *subpacificus* in the surface layer, *Thalassiosira* sp. (180 cells l<sup>-1</sup>) at 25 m depth and *Dactyliosolen* sp. at 60 m depth were encountered (Table 2).

## 4 Discussion

### 4.1 High Nutrient-Low Chlorophyll phytoplankton assemblages

The HNLC-PA in the southeast Pacific was dominated by the lightly silicified diatoms *Pseudo-nitzschia delicatissima* and *Rhizosolenia bergonii* that were encountered in the Marquesas Islands Archipelago at 8° S with a surface temperature of 28° C and salinity 35.5, and between 2000 and 600 km off the Chilean coast at 33° S with a surface temperature of 17° C and salinity of 34 (Figs. 2–3, 8–9). The western side of the PCC did not appear in the literature as one of the HNLC regions of the world's oceans. The HNLC-PA showed scarce differences in these geographically distant regions. Only a few tropical species such as the dinoflagellate genus

*Citharistes* was absent in temperate waters (Table 2). HNLC diatoms such as *Planktoniella sol* or *Rhizosolenia bergonii* are commonly reported along the South American coastal waters associated with warm waters especially during the El Niño conditions (Avaria and Muñoz, 1987; Rodríguez et al., 1996). Based on a single survey and with a paucity of information in the literature in offshore waters in the region, the present study does not allow to establish the occurrence of the HNLC-PA in the western PCC as a permanent feature. In the SE Pacific the HNLC-PA was encountered in two regions of contrasted temperature and salinity values. Other trophic or ecological factor(s) may determinate the occurrence of the HNLC-PA in these two distant regions.

#### 4.1.1 Grazing and the HNLC-PA

One of the explanations for the HNLC paradox is that phytoplankton populations are strictly controlled by the zooplankton (Frost and Franzen, 1992; Leising et al., 2003). In the equatorial Pacific, Roman and Gauzens (1997) suggested that the scarce copepods mainly grazed on protozoa. Landry et al. (1995) reported that grazing by microzooplankton dominates phytoplankton losses, and that large diatoms are not easily exploited by protistan consumers (ciliates, dinoflagellates, etc). The microscope observations in the present study showed a very low abundance of copepods, although the scarce volume examined (0.5 l) were insufficient to provide a statistically valid estimation of the early stages of copepods abundance. The ciliates and heterotrophic dinoflagellates (*Gyrodinium* spp. and *Protoperidinium* spp.) were abundant.

The two most successful species in the HNLC regions of the SE Pacific, *Pseudo-nitzschia delicatissima* and *Rhizosolenia bergonii*, seems to develop two different anti-grazing strategies. *Pseudo-nitzschia delicatissima*, and occasionally *Nitzschia longissima* increased their sizes by forming clumps or “ball of needles” of  $\sim 100\ \mu\text{m}$  in diameter disposed in all directions (Figs. 10–11). This diatom behavior was already described by Hasle (1960) and Buck and Chavez (1994). The aggregation of these diatoms suggests an anti-grazing adaptation in an environment dominated by micrograzers. On the other hand, centric diatoms such as *Rhizosolenia bergonii* and *R. acuminata* reached sizes  $> 500\ \mu\text{m}$  in length, being far of the prey size spectra of most micrograzers.

Other diatom species are not exclusive of the HNLC conditions, although they are favoured in these eutrophic conditions. Diatoms showed strongly elongated frustules or extensions. For example *Thalassiothrix longissima* is thin, but reached  $2000\ \mu\text{m}$  in length. *Planktoniella sol* secretes a large hyaline ring that increases its size. *Chaetoceros peruvianus* is a uni-cellular species with long spine-bearing setae. Colonial species such as *Chaetoceros atlanticus* var. *neapolianus* and *Bacteriastrium* cf. *elongatum* showed a similar strategy with long setae.

Beyond the size increase or aggregation of the diatoms as possible response to small grazers, it should be taken into account that the assemblage was dominated by *Pseudo-nitzschia*, a genus that contains numerous species able to produce the domoic acid. Reactive aldehydes produced by diatoms have been reported to reduce the fecundity and egg hatching success of copepods (Ianora et al., 2003). The effects of the compounds produced by diatoms such as the domoic acid, if produced, on the abundance of copepods in the HNLC region need further research.

#### 4.1.2 Lightly silicified diatoms near the surface?

A rigid and thick frustule of diatoms is considered to be the main anti-grazing strategy against copepods. If copepods are not abundant in HNLC conditions, lightly silicified diatoms may be favoured. The dominant diatoms of the HNLC regions are characterized by thin frustules and the lack siliceous ornamentation requiring a superfluous silicon uptake. *Pseudo-nitzschia delicatissima* and *R. bergonii* easily dissolved their lightly silicified frustules and these diatoms are nearly absent from the sediments (Kolbe, 1954). At first sight, a lightly silicified frustule may be a response to an environment where the silicic acid bioavailability is low. In the present study the HNLC-PA appeared in two regions with different hydrographical conditions, but apparent similar trophic conditions. The *P. delicatissima*-*R. bergonii* assemblage (Figs. 8–9) was associated with a nitrate-silicic acid ratio that ranged from 1–3 in the euphotic zone of the two HNLC regions (Fig. 6, Table 1). The elemental composition of the diatoms is characterized by nitrogen-silicic acid ratio of 1 (Brzezinski, 1985). It can be expected that the diatoms uptake the nutrients from the surrounding waters with a similar ratio. Values of the nitrate-silicic acid ratio higher than 1, may imply a silicon limitation for the diatom growth. However, the nutrient limitation is a complex issue, and only the nutrient ratios are not suitable to infer limitations. Based on silicic acid uptake measurements, Leynaert et al. (2001) reported direct evidence that the diatoms are Si-limited in the central equatorial Pacific.

It is evident that the diatoms must be adapted to optimize the silicic acid utilization. Large frustules such as those in *R. bergonii* and *R. acuminata* may allow to escape of most of micrograzers. The balance between large frustules avoiding the micrograzers and a thin frustule that allowed to economize the scarce silicic acid available may result in deficiencies in the formation and maintenance of the frustule. In the present study, specimens of *Rhizosolenia bergonii* and *R. acuminata* were curved instead of the usual rigid frustules and sections of the frustules appeared anomalously fragmented (Figs. 12–15). This phenomenon did not occur with other diatoms and the plasmalemma of the diatom appeared retracted in the regions where the frustule is fragmented. The portion of the frustule closer to the diatom nucleus did not usually appear fragmented (Figs. 14–15). Consequently this

phenomenon of fragmentation of the frustule occurred before the fixation of the samples, discarding an artifact due to the fixation methodology. This is evidence that the silicic acid is an element limiting the large diatoms *R. bergonii* and *R. acuminata* in the HNLC regions of the South Pacific Ocean.

The possible deficiency of silicic acid may determinate the presence of other phytoplankton components of the HNLC-PA. Chavez (1989) included the absence of neritic bloom-forming diatoms among the factors to explain the HNLC paradox. Although colonial diatoms such as *Chaetoceros lorenzianus* were observed in the present study, well silicified neritic diatoms (i.e. *Detonula*, *Leptocylindrus*, *Guinardia*, etc.) that are dominant in mesotrophic coastal waters were absent in the HNLC regions. The typical neritic diatoms may be less competitive in the HNLC regions versus the lightly silicified diatoms. For example *Planktoniella sol* has developed an organic ring-like wing that does not contain silica. Beyond the diatoms, other phytoplankton groups such as silicoflagellates have an internal silica skeleton. *Dityocha* is often a common component of the phytoplankton in the north Pacific Gyre (Venrick, 1992; Scharek et al., 1999), were nearly absent during this survey in the SE Pacific. The open SE Pacific is far from the terrestrial inputs of dissolved silicon. The Antarctic waters spread along the deep SE Pacific Ocean. The waters of the PCC showed a nitrate-silicic acid ratio higher than 1 that may be associated to a Si-deficiency (Fig. 6). At the bottom of the euphotic zone the nitrate-silicic acid ratio was also higher than 1 (Fig. 6). Compared to the other oceanic regions, the SE Pacific seems to show deficiencies of the stock of silicic acid compared to other major nutrients. However, it is uncertain whether silicic acid is the main factor that determinate the dominant HNLC-PA and the diatoms of the bottom of the euphotic zone in the SPG.

Leynaert et al. (2001) hypothesized that silicon and iron limitations may interact. Iron is an essential component for the synthesis of chlorophyll and the nitrate reductase of the primary producers. It is unknown whether the iron from the Marquesas Islands Archipelago is the main responsible of the increase of the abundance of *Pseudo-nitzschia* and/or the increase is mainly due to the entrainment of deep major nutrients due the “island mass effect” (Signorini et al., 1999). In the present study, offshore the Marquesas Islands Archipelago, the dominance of the small *Pseudo-nitzschia* decreased and larger centric diatoms such as *R. bergonii* showed higher abundance (Fig. 8). In the proximity of islands the major nutrients and iron inputs is expected to be more continuous, favouring smaller diatoms such as *Pseudo-nitzschia*. In offshore waters, the pulsating enrichment events may favour species with big vacuoles such as *R. bergonii* able to accumulate nutrients for oligotrophic periods. Several studies showed that the domoic acid produced by *Pseudo-nitzschia* binds iron and copper (Rue and Bruland, 2001; Wells et al., 2005). The domoic acid is a strong organic ligand that facilitates the iron uptake of *Pseudo-nitzschia*

spp. According to Wells et al. (2005) this may explain why *Pseudo-nitzschia* spp. have persistent populations in oceanic HNLC regions.

Other characteristic of the HNLC diatoms is that they are restricted to surface waters and they did not form the typical sub-surface maximum as usual in other diatoms. For example species considered as deep flora such as *Planktoniella sol* (Sournia, 1982) showed a surface distribution in the HNLC regions of the present study. Under unfavourable surface conditions the diatoms are expected to sink to deep waters. However, in the transition of the warm HNLC region to oligotrophic conditions (13°32' S–132°07' W) *Rhizosolenia bergonii* remained near the surface despite the nitrate stock was depleted (Figs. 7–8, Table 1). It is uncertain the factor that determinate the surface distribution of these diatoms. Near the surface, the irradiance is higher and atmospheric inputs of iron are available. Iron is required for the photosynthetic electron transport system and chlorophyll molecule biosynthesis. At deeper waters (low irradiance), it can be expected that diatoms will require the synthesis of more chlorophyll and electron transport system molecules. Several studies reported that the iron requirements for diatoms increase at sub-saturating irradiances and decrease the efficiency of the utilization of iron (Sunda and Huntsman, 1995, 1997; Muggli and Harrison, 1997). Consequently in iron-depleted waters, the diatoms near the surface may grow with less iron requirements than in deeper waters. The hypothesis that the iron-stress would determinate the vertical distribution of HNLC diatoms requires further research. Other features such as the anti-grazing strategies and the physiology of HNLC species *Pseudo-nitzschia delicatissima* and *Rhizosolenia bergonii* need to be investigated.

Environmental factors, other than temperature and salinity, are responsible for the establishment and the persistence of a weakly silicified diatom assemblage in two geographically and hydrologically distant regions, the tropical central Pacific and the western limit of the Perú-Chile Current during the austral summer. The nitrate-silicic acid ratio ranging from 1–3 in the euphotic zone and frustules anomalously fragmented in *Rhizosolenia bergonii* suggest that Si-deficiency may be responsible of the HNLC-PA. Further studies are required to establish the factors that favour the occurrence of this distinctive phytoplankton assemblage.

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